

# Package ‘BEAMR’

January 20, 2025

**Title** Bootstrap Evaluation of Association Matrices

**Version** 1.1.0

**Description** A bootstrap-based approach to integrate multiple forms of high dimensional genomic data with multiple clinical endpoints. This method is used to find clinically meaningful groups of genomic features, such as genes or pathways. A manuscript describing this method is in preparation.

**License** GPL (>= 3)

**Encoding** UTF-8

**RoxygenNote** 7.3.2

**Imports** dplyr, ggmosaic, ggplot2, ggpubr, logistf, magrittr, MASS, purrr, rlist, stats, stringr, survival, survminer

**Suggests** rmarkdown

**Depends** R (>= 2.10)

**LazyData** true

**URL** <https://annaseffernick.github.io/BEAMR/>,  
<https://github.com/annaSeffernick/BEAMR>

**BugReports** <https://github.com/annaSeffernick/BEAMR/issues>

**NeedsCompilation** no

**Author** Anna Eames Seffernick [aut, cre, cph]  
(<<https://orcid.org/0000-0003-0848-4604>>),  
Stanley Pounds [aut],  
Xueyuan Cao [aut]

**Maintainer** Anna Eames Seffernick <[anna.seffernick@stjude.org](mailto:anna.seffernick@stjude.org)>

**Repository** CRAN

**Date/Publication** 2024-07-27 16:00:06 UTC

## Contents

beam_dat . . . . .	2
beam_dat_sm . . . . .	3
beam_specs . . . . .	4
beam_specs_sm . . . . .	4
beam_stats . . . . .	5
beam_stats_sm . . . . .	5
check_beam_specs . . . . .	6
check_list_class . . . . .	6
clean_Bmtx . . . . .	7
clinf . . . . .	7
compute_beam_stats . . . . .	8
compute_feature_pvalues . . . . .	9
compute_set_pvalues . . . . .	9
extend_set_data . . . . .	10
extract_beam_stats . . . . .	11
find_id_clm . . . . .	11
gen_beam_plot_list . . . . .	12
get_id_index . . . . .	13
omicann . . . . .	14
omicdat . . . . .	14
plot_beam_boot . . . . .	15
plot_beam_clin . . . . .	16
plot_feat_clin . . . . .	17
prep_beam_data . . . . .	18
prep_beam_plot . . . . .	19
prep_beam_specs . . . . .	20
print.beam.data . . . . .	21
print.beam.stats . . . . .	22
setdat . . . . .	22
specs . . . . .	23
subset_beam_result . . . . .	23
<b>Index</b>	<b>26</b>

---

 beam\_dat

*Pediatric T-ALL Clinical Data from COG trial AALL0434*


---

### Description

The beam.data object used in example beam analyses

### Usage

beam\_dat

**Format**

beam\_dat:

A beam.data object, which is a list with the following elements:

**main.data** A data.frame with clinical/endpoint data.

**mtx.data** A list of the omics data matrices.

**mtx.anns** A list of omic annotation data.frames.

**anns.mtch** A data.frame with information to link mtx.data and mtx.anns.

**set.data** A data.frame with set.id, mtx.id, and row.id to link omic features to sets.

**set.anns** Optional data.frame with set annotation data.

**boot.index** A matrix with bootstrap indices.

**Source**

NA

---

beam\_dat\_sm

*Pediatric T-ALL Clinical Data from COG trial AALL0434*

---

**Description**

The smaller beam.data object used in the example for compute\_beam\_stats function

**Usage**

beam\_dat\_sm

**Format**

beam\_dat\_sm:

A beam.data object, which is a list with the following elements:

**main.data** A data.frame with clinical/endpoint data.

**mtx.data** A list of the omics data matrices.

**mtx.anns** A list of omic annotation data.frames.

**anns.mtch** A data.frame with information to link mtx.data and mtx.anns.

**set.data** A data.frame with set.id, mtx.id, and row.id to link omic features to sets.

**set.anns** Optional data.frame with set annotation data.

**boot.index** A matrix with bootstrap indices.

**Source**

NA

---

beam_specs	<i>Pediatric T-ALL BEAM Analysis Specs Data from COG trial AALL0434</i>
------------	---

---

**Description**

The beam.specs object used in example beam analyses

**Usage**

```
beam_specs
```

**Format**

beam\_specs:

A data frame with 6 rows and 3 columns:

**name** Analysis name with omic and endpoint

**mtx** Name of omics matrix used in the analysis

**mdl** Regression model

**Source**

NA

---

beam_specs_sm	<i>Pediatric T-ALL BEAM Analysis Specs Data from COG trial AALL0434</i>
---------------	---

---

**Description**

The small beam.specs object used in example compute\_beam\_stats function.

**Usage**

```
beam_specs_sm
```

**Format**

beam\_specs\_sm:

A data frame with 2 rows and 3 columns:

**name** Analysis name with omic and endpoint

**mtx** Name of omics matrix used in the analysis

**mdl** Regression model

**Source**

NA

---

beam_stats	<i>Pediatric T-ALL Clinical Data from COG trial AALL0434</i>
------------	--

---

**Description**

The beam.stats object used in example beam analyses

**Usage**

```
beam_stats
```

**Format**

beam\_stats:

A beam.stats object, which contains the following objects

**beam.stats** A list of data.frames of association statistics for each omic-endpoint pair.

**beam.specs** A beam.specs object (data.frame with name, mtx, and mdl.)

**beam.data** The beam.data object.

**Source**

NA

---

beam_stats_sm	<i>Pediatric T-ALL Clinical Data from COG trial AALL0434</i>
---------------	--

---

**Description**

The small beam.stats object used in example for compute\_beam\_stats function.

**Usage**

```
beam_stats_sm
```

**Format**

beam\_stats\_sm:

A beam.stats object, which contains the following objects

**beam.stats** A list of data.frames of association statistics for each omic-endpoint pair.

**beam.specs** A beam.specs object (data.frame with name, mtx, and mdl.)

**beam.data** The beam.data object.

**Source**

NA

---

check_beam_specs	<i>Check that beam.specs satisfies all necessary conditions</i>
------------------	---

---

**Description**

Check that beam.specs satisfies all necessary conditions

**Usage**

```
check_beam_specs(beam.specs, mtx.names)
```

**Arguments**

beam.specs	A data.frame with column name, mtx, and mdl
mtx.names	A vector with the names of the data matrices (beam.data\$mtx.data)

**Value**

A data.frame of beam.specs if all conditions satisfied, otherwise throws an error

**Examples**

```
data(beam_dat)
data(beam_specs)
test_specs <- check_beam_specs(beam_specs, names(beam_dat$mtx.data))
```

---

check_list_class	<i>Check that each element of a list is of a required class</i>
------------------	---

---

**Description**

Check that each element of a list is of a required class

**Usage**

```
check_list_class(list.object, required.class)
```

**Arguments**

list.object	A list used in BEAMR analysis
required.class	Class for list elements, e.g. matrix

**Value**

Logical TRUE if list is of required class

**Examples**

```
data(omicdat)
check_list_class(omicdat, "matrix")
```

---

clean_Bmtx	<i>Clean up bootstrap coefficient matrix</i>
------------	--

---

**Description**

Clean up bootstrap coefficient matrix

**Usage**

```
clean_Bmtx(B)
```

**Arguments**

B                    Matrix of bootstrap coefficients

**Value**

Matrix of cleaned bootstrap coefficients

**Examples**

```
data(beam_stats)
B.mtx <- beam_stats$beam_stats[[1]]
B.cln <- clean_Bmtx(B.mtx)
```

---

clinf	<i>Pediatric T-ALL Clinical Data from COG trial AALL0434</i>
-------	--

---

**Description**

A subset of clinical data from pediatric and young adult t-lineage acute lymphoblastic leukemia patients in the Children's Oncology Group trial AALL0434, published in Liu et al., 2017 Nature Genetics

**Usage**

```
clinf
```

**Format**

**clinf:**  
 A data frame with 265 rows and 8 columns:  
**ID** Subject ID  
**MRD29** Minimal residual disease measured at day 29  
**RNA.clm** Key to match to RNA matrix  
**Lesion.clm** Key to match Lesion matrix  
**Lesion.id** Key to match Lesion matrix  
**RNA.id** Key to match RNA matrix  
**EFS** Event-free survival Surv object  
**OS** Overall survival Surv object

**Source**

<https://www.nature.com/articles/ng.3909>

---

compute\_beam\_stats      *Compute bootstrap model coefficients for BEAM*

---

**Description**

Compute bootstrap model coefficients for BEAM

**Usage**

```
compute_beam_stats(beam.data, beam.specs, stdize = TRUE)
```

**Arguments**

beam.data	Result of prep.beam.data
beam.specs	A data.frame of strings with columns name, mtx, mdl (string with R model with mtx.row)
stdize	Logical whether to standardize (center and scale) predictors or not. Default is TRUE.

**Value**

A beam.stats object, which is a list with beam.stats (the association matrices), the beam.specs, and the beam.data

**Examples**

```

data(beam_dat_sm)
data(beam_specs_sm)
test.beam.stats <- compute_beam_stats(beam.data=beam_dat_sm,
                                     beam.specs=beam_specs_sm, stdize=TRUE)

```

---

`compute_feature_pvalues`*Compute feature level p-values from BEAM statistics*

---

**Description**

Compute feature level p-values from BEAM statistics

**Usage**

```
compute_feature_pvalues(beam.stats)
```

**Arguments**

`beam.stats` A beam.stats object, which is a list with beam.stats (the association matrices), the beam.specs, and the beam.data

**Value**

A list of feature level p-values, with each entry a data frame for a different omics/endpoint association, with columns id, gene, beta, p, q

**Examples**

```
data(beam_stats)
test.featt.pvals <- compute_feature_pvalues(beam.stats=beam_stats)
```

---

`compute_set_pvalues` *Compute BEAMR p-values for sets*

---

**Description**

Compute BEAMR p-values for sets

**Usage**

```
compute_set_pvalues(
  beam.stats,
  peel = FALSE,
  z = TRUE,
  alpha = 0.1,
  mess.freq = 25
)
```

**Arguments**

beam.stats	A beam.stats object from compute_beam_stats function
peel	Logical indicating whether to peel in p-value calculation
z	Logical indicating whether to z-scale each vector of one coefficient estimate across bootstraps before analysis
alpha	Maximum depth to peel (reduces computing time); default 0.1.
mess.freq	Message frequency; default 25.

**Value**

A list with a data.frame of set p-values from BEAMR analysis, a data.frame of summary row p-values, and a data frame of set matching.

**Examples**

```
data(beam_stats_sm)
test.pvals <- compute_set_pvalues(beam.stats=beam_stats_sm)
```

---

extend_set_data	<i>Extend set definition data with genes on the same row separated by commas, semicolons, slashes, etc</i>
-----------------	--

---

**Description**

Extend set definition data with genes on the same row separated by commas, semicolons, slashes, etc

**Usage**

```
extend_set_data(set.data, sep)
```

**Arguments**

set.data	A data frame with set definition data.
sep	Punctuation to split on.

**Value**

A data frame.

**Examples**

```
data(setdat)
extend_set_data(setdat, sep=",")
```

---

extract_beam_stats	<i>Extract beam stats for a specific set</i>
--------------------	--

---

**Description**

Extract beam stats for a specific set

**Usage**

```
extract_beam_stats(beam.stats, set.id)
```

**Arguments**

beam.stats	A beam.stats object, which is a list with beam.stats (the association matrices), the beam.specs, and the beam.data
set.id	A character of a set id name (an entry in in beam.data\$set.data\$set.id)

**Value**

A matrix with with estimated associations for each endpoint and each omic feature linked to the set

**Examples**

```
data(beam_stats)
test.stats <- extract_beam_stats(beam_stats, set.id="ENSG00000099810")
```

---

find_id_clm	<i>Find the column of mtch.data with the most rows containing an element of ids</i>
-------------	---

---

**Description**

Find the column of mtch.data with the most rows containing an element of ids

**Usage**

```
find_id_clm(mtch.data, ids)
```

**Arguments**

mtch.data	A data.frame
ids	A vector of row ids to match

**Value**

A vector of column names with the most matches.

**Examples**

```

data(omicann)
data(omicdat)
lsn.data <- omicann[[1]]
mtx.rows <- rownames(omicdat[[1]])
test <- find_id_clm(lsn.data,mtx.rows)

```

---

gen\_beam\_plot\_list      *Generate BEAM Plot List*

---

**Description**

Internal function: generate a list of clinical feature plots.

**Usage**

```

gen_beam_plot_list(
  beam.result,
  beam.specs,
  beam.featt.pvals,
  number.pairs = 1,
  set.id,
  feat.id = NULL,
  title.size = 10,
  pair.order = "both",
  endpt.order = NULL
)

```

**Arguments**

beam.result	Result of prep.beam.data
beam.specs	A data.frame of strings with columns name, mtx, mdl, plot
beam.featt.pvals	List of feature-level p-values from compute_feature_pvalues
number.pairs	Numeric; number of features to display in clinical plots, ordered by significance
set.id	A character with set name; must be in beam.result\$beam.data\$set.data\$set.id
feat.id	Default NULL; a character with feature name; must be in beam.result\$beam.data\$set.data\$row.id
title.size	A numeric. Specify the size of individual plot titles. Default is 10.
pair.order	One of c("both", "omic", "endpoint"). Default is "both." Specify how to choose feature-endpoint plots to include. If "both", find the best (based on q, p, effect size) feature-omic pair for each type of omic and each endpoint separately. If "omic", within each omic, find the best feature-endpoint pair and then plot this feature with all endpoints. If "endpoint", need to specify endpt.order as the name of chosen endpoint. Then, within each omic, find the feature with best association with the selected endpoint, and plot this feature for all endpoints.
endpt.order	Default NULL. If pair.order="endpoint", specify character with endpoint name (from beam.specs\$name, after the period).

**Value**

A list of plots for the specified set and/or feature.

**Examples**

```
data(beam_stats)
test.featt.pvals <- compute_feature_pvalues(beam.stats=beam_stats)
plot.specs <- prep_beam_plot(beam.data=beam_stats$beam.data,
                             beam.specs=beam_stats$beam.specs)
plot.list <- gen_beam_plot_list(beam.result=beam_stats, beam.specs=plot.specs,
                                beam.featt.pvals=test.featt.pvals,
                                number.pairs=1, set.id="ENSG00000099810",
                                feat.id=NULL, title.size=11,
                                pair.order="omic", endpt.order=NULL)
```

---

get_id_index	<i>For each row of the data.frame main.data, find the index of the matching element in vector ids</i>
--------------	---

---

**Description**

For each row of the data.frame main.data, find the index of the matching element in vector ids

**Usage**

```
get_id_index(mtch.data, ids, warn = TRUE)
```

**Arguments**

mtch.data	A data.frame to be linked with the ids
ids	A vector of ids to be linked in mtch.data
warn	A logical value whether to include warnings with results

**Value**

A data.frame with matching id index

**Examples**

```
data(clinf)
data(omicdat)
mtx.clms <- colnames(omicdat[[1]])
id_index <- get_id_index(clinf,mtx.clms)
```

---

`omicann`*Pediatric T-ALL Omics Annotation Data from COG trial AALL0434*

---

**Description**

A subset of genomic lesion and RNA expression data from pediatric and young adult t-lineage acute lymphoblastic leukemia patients in the Children's Oncology Group trial AALL0434, published in Liu et al., 2017 Nature Genetics. This is the annotation mapping feature id to gene name given by Ensembl ID.

**Usage**`omicann`**Format**`omicann:`

A list with two data frames of omics annotation.

**Lesion** A dataframe with 20 rows and 2 columns with lesion ID and Ensembl ID.

**RNA** A dataframe with 20 rows and 2 columns with feature ID and Ensembl ID.

**Source**

<https://www.nature.com/articles/ng.3909>

---

`omicdat`*Pediatric T-ALL Omics Data from COG trial AALL0434*

---

**Description**

A subset of genomic lesion and RNA expression data from pediatric and young adult t-lineage acute lymphoblastic leukemia patients in the Children's Oncology Group trial AALL0434, published in Liu et al., 2017 Nature Genetics

**Usage**`omicdat`**Format**`omicdat:`

A list with two dataframes of omic data for each subject

**Lesion** A dataframe with 20 rows and 265 columns indicating presence of lesion.

**RNA** A dataframe with 20 rows and 265 columns with expression data.



---

plot_beam_clin	<i>Plot BEAM Sets</i>
----------------	-----------------------

---

### Description

plot\_beam\_clin produces a matrix of feature level clinical plots for a set. Users can specify which omic/endpoint pairs they want to see as well as the number of features from the set. Default is all omic/endpoint pairs and the top feature (smallest feature-level p-value).

### Usage

```
plot_beam_clin(
  beam.result,
  beam.specs = NULL,
  beam.set.pvals,
  beam.feats.pvals,
  set.id,
  gene.name = NULL,
  pair.type = NULL,
  number.pairs = 1,
  pair.order = "both",
  endpt.order = NULL,
  n.col = NULL,
  n.row = NULL,
  title.size = 10
)
```

### Arguments

beam.result	A beam.stats object from compute_beam_stats
beam.specs	A data.frame. Default NULL, in which case beam.result\$beam.specs is used. Otherwise can input other beam.specs data.frame that must contain name, mtx, mdl, plot columns.
beam.set.pvals	A list containing BEAMR set p-values from compute_set_pvalues.
beam.feats.pvals	A list containing feature-level p-values from compute_feature_pvalues.
set.id	A character specifying the name of a set. Must be in beam.result\$beam.data\$set.data
gene.name	A character specifying a Gene Name/Symbol for the set. Default is NULL
pair.type	A character vector. Default NULL, in which case clinical plots for all omic/endpoint pairs are produced. Otherwise specify pairs from beam.stats\$beam.specs\$name
number.pairs	A numeric. Default 1, in which case only feature with best simple test for each pair is plotted. If >1, show top n simple plots ordered by feature-level p-value
pair.order	One of c("both", "omic", "endpoint"). Default is "both." Specify how to choose feature-endpoint plots to include. If "both", find the best (based on q, p, effect

size) feature-omic pair for each type of omic and each endpoint separately. If "omic", within each omic, find the best feature-endpoint pair and then plot this feature with all endpoints. If "endpoint", need to specify endpt.order as the name of chosen endpoint. Then, within each omic, find the feature with best association with the selected endpoint, and plot this feature for all endpoints.

endpt.order	Default NULL. If pair.order="endpoint", specify character with endpoint name (from beam.specs\$name, after the period).
n.col	A numeric. Specify the number of columns for the plot layout; default NULL will use the number of omics types.
n.row	A numeric. Specify the number of rows for the plot layout; default NULL will automatically define the number of rows after number of columns specified.
title.size	A numeric. Specify the size of individual plot titles. Default is 10.

**Value**

A figure (ggarrange object)

**Examples**

```
data(beam_stats)
test.pvals <- compute_set_pvalues(beam.stats=beam_stats)
test.featt.pvals <- compute_feature_pvalues(beam.stats=beam_stats)
plot.specs <- prep_beam_plot(beam.data=beam_stats$beam.data,
                             beam.specs=beam_stats$beam.specs)
test.plot <- plot_beam_clin(beam.result=beam_stats, beam.specs=plot.specs,
                            beam.set.pvals=test.pvals,
                            beam.featt.pvals=test.featt.pvals,
                            set.id="ENSG00000099810", gene.name="MTAP",
                            pair.type=NULL, number.pairs=1, n.col=4,
                            n.row=NULL, title.size=11,
                            pair.order="omic", endpt.order=NULL)
```

---

plot\_feat\_clin

*Plot BEAM Feature*

---

**Description**

plot\_feat\_clin produces a matrix of feature level clinical plots for a specific feature.

**Usage**

```
plot_feat_clin(
  feat.id,
  beam.result,
  beam.specs = NULL,
  beam.set.pvals,
  beam.featt.pvals,
```

```

    n.row = NULL,
    n.col = NULL
  )

```

### Arguments

`feat.id` A character specifying the name of a feature. Must be in `beam.result$beam.data$set.data`

`beam.result` A `beam.stats` object from `compute_beam_stats`

`beam.specs` A `data.frame`. Default `NULL`, in which case `beam.result$beam.specs` is used. Otherwise can input other `beam.specs` `data.frame` that must contain `name`, `mtx`, `mdl`, `plot` columns.

`beam.set.pvals` A list containing BEAMR set p-values from `compute_set_pvalues`.

`beam.featt.pvals` A list containing feature-level p-values from `compute_feature_pvalues`.

`n.row` A numeric. Specify the number of rows for the plot layout; default `NULL` will automatically define the number of rows after number of columns specified.

`n.col` A numeric. Specify the number of columns for the plot layout; default `NULL` will use the number of omics types.

### Value

A figure (ggarrange object)

### Examples

```

data(beam_stats)
test.pvals <- compute_set_pvalues(beam.stats=beam_stats)
test.featt.pvals <- compute_feature_pvalues(beam.stats=beam_stats)
plot.specs <- prep_beam_plot(beam.data=beam_stats$beam.data, beam.specs=beam_stats$beam.specs)
test.plot <- plot_feat_clin(beam.result=beam_stats, beam.specs=plot.specs,
                           beam.set.pvals=test.pvals, beam.featt.pvals=test.featt.pvals,
                           feat.id="ENSG00000227443_loss",
                           n.col=2, n.row=NULL)

```

---

```
prep_beam_data
```

*Prepare data for BEAM analysis*

---

### Description

Prepare data for BEAM analysis

**Usage**

```

prep_beam_data(
  main.data,
  mtx.data,
  mtx.anns = NULL,
  set.data = NULL,
  set.anns = NULL,
  n.boot = 1000,
  seed = NULL
)

```

**Arguments**

main.data	A data.frame
mtx.data	A list, each element is a matrix
mtx.anns	A list, each element is a data.frame
set.data	A data.frame with columns set.id, mtx.id, row.id
set.anns	A data frame with set.id and other columns
n.boot	Number of bootstraps
seed	Initial seed for random number generation

**Value**

A beam.data object, which is a list with main.data, mtx.data, mtx.anns, anns.mtch, set.data, set.anns, and boot.index

**Examples**

```

data(clinf)
data(omicdat)
data(omicann)
data(setdat)
test.beam.data <- prep_beam_data(main.data=clinf, mtx.data=omicdat,
                                mtx.anns=omicann, set.data=setdat,
                                set.anns=NULL, n.boot=10, seed=123)

```

---

```

prep_beam_plot      Prepare for BEAM plotting

```

---

**Description**

Add a "plot" column to beam.specs, which includes string of plot commands.

**Usage**

```

prep_beam_plot(beam.data, beam.specs)

```

**Arguments**

beam.data	Result of prep.beam.data
beam.specs	A data.frame of strings with columns name, mtx, mdl (string with R model with mtx.row)

**Value**

An updated beam.specs object that includes the column "plot"

**Examples**

```
data(clinf)
data(omicdat)
data(omicann)
data(setdat)
test.beam.data <- prep_beam_data(main.data=clinf, mtx.data=omicdat,
                                mtx.anns=omicann, set.data=setdat,
                                set.anns=NULL, n.boot=10, seed=123)
specs <- prep_beam_specs(beam.data=test.beam.data, endpts=c("MRD29", "EFS", "OS"),
                        firth=TRUE)
plot.specs <- prep_beam_plot(beam.data=test.beam.data, beam.specs=specs)
```

---

```
prep_beam_specs      Prepare beam.specs
```

---

**Description**

Prepare the beam.specs data.frame for BEAM model fitting. Specifies the univariate models needed to compute the BEAMR set p-values.

**Usage**

```
prep_beam_specs(
  beam.data,
  endpts,
  firth = TRUE,
  adjvars = NULL,
  endptmdl = NULL
)
```

**Arguments**

beam.data	A beam.data object from prep_beam_data
endpts	A vector of endpoint variable names in main.data
firth	A logical value. If TRUE (default) fit Firth penalized Cox model to account for monotone likelihood in the presence of rare events or predictors. If FALSE fit usual Cox model.

adjvars	Default NULL, optional vector of adjustment variable names in main.data
endptmdl	Optional model specification data.frame with endpoint name column called "endpt" and model string column called "mdl"

**Value**

The beam.specs object, a data.frame specifying the omics-endpoint association models to be fit

**Examples**

```
data(clinf)
data(omicdat)
data(omicann)
data(setdat)
test.beam.data <- prep_beam_data(main.data=clinf, mtx.data=omicdat,
                                mtx.anns=omicann, set.data=setdat,
                                set.anns=NULL, n.boot=10, seed=123)

#Without adjustment
prep_beam_specs(beam.data=test.beam.data, endpts=c("MRD29", "OS", "EFS"),
                firth=TRUE)
# With adjustment
prep_beam_specs(beam.data=test.beam.data, endpts=c("OS", "EFS"),
                adjvars=c("MRD29"), firth=TRUE)
```

---

```
print.beam.data      Print summary information about a beam.data object
```

---

**Description**

Print summary information about a beam.data object

**Usage**

```
## S3 method for class 'beam.data'
print(x, ...)
```

**Arguments**

x	An object of class "beam.data"
...	Other arguments passed to or from other methods

**Value**

Messages about the beam.data object

**Examples**

```
data(beam_dat)
print(beam_dat)
```

---

```
print.beam.stats      Print summary information about beam.stats object
```

---

### Description

Print summary information about beam.stats object

### Usage

```
## S3 method for class 'beam.stats'
print(x, ...)
```

### Arguments

```
x          An object of class "beam.stats"
...        Other arguments passed to or from other methods
```

### Value

Messages about the beam.data object

### Examples

```
data(beam_stats)
print(beam_stats)
```

---

```
setdat      Map of Pediatric Data from COG trial AALL0434
```

---

### Description

Map between annotation and omic data for a subset of clinical data from pediatric and young adult t-lineage acute lymphoblastic leukemia patients in the Children's Oncology Group trial AALL0434, published in Liu et al., 2017 Nature Genetics

### Usage

```
setdat
```

### Format

**setdat:**

A data frame with 40 rows and 3 columns

**set.id** Ensembl ID that defines gene-feature set

**mtx.id** Name of omic matrix where corresponding feature data can be found

**row.id** Feature name in corresponding omic matrix

**Source**

<https://www.nature.com/articles/ng.3909>

---

specs	<i>Pediatric T-ALL BEAMR Analysis Specs Data from COG trial AALL0434</i>
-------	--

---

**Description**

The beam.specs object used in example beam analyses

**Usage**

```
specs
```

**Format**

specs:

A data frame with 6 rows and 3 columns:

**name** Analysis name with omic and endpoint

**mtx** Name of omics matrix used in the analysis

**mdl** Regression model

**Source**

NA

---

subset_beam_result	<i>Subset beam.stats Result</i>
--------------------	---------------------------------

---

**Description**

Filter the beam.stats object from compute\_beam\_stats with various filtering criteria. Default is to filter to top 50 sets with smallest q-value. At least one filtering criteria must be specified. Can also use intersection or union of multiple criteria.

**Usage**

```
subset_beam_result(
  beam.result,
  beam.set.pvals = NULL,
  beam.featt.pvals = NULL,
  mtx.rows = NULL,
  set.ids = NULL,
  endpts = NULL,
  omics = NULL,
  p.limit = NULL,
  q.limit = NULL,
  p.featt.limit = NULL,
  q.featt.limit = NULL,
  intersect = TRUE,
  recalc = FALSE
)
```

**Arguments**

<code>beam.result</code>	A <code>beam.stats</code> object from <code>compute_beam_stats</code>
<code>beam.set.pvals</code>	A list containing BEAMR set p-values from <code>compute_set_pvalues</code> ; required if <code>p.limit</code> or <code>q.limit</code> are specified.
<code>beam.featt.pvals</code>	A list containing feature-level p-values from <code>compute_feature_pvalues</code> ; required if <code>p.featt.limit</code> or <code>q.featt.limit</code> are specified.
<code>mtx.rows</code>	A list of vectors of feature names corresponding to <code>row.id</code> in <code>set.data</code> . List names correspond to <code>mtx.id</code> in <code>set.data</code> . If specified, filter to all sets containing at least one of these features.
<code>set.ids</code>	A character vector of <code>set.ids</code> . If specified, filter to these sets.
<code>endpts</code>	A character vector of endpoint names. If specified, filter to sets that correspond to these endpoints.
<code>omics</code>	A character vector of omics names. If specified, filter to sets that correspond to these omics.
<code>p.limit</code>	A numeric value. If specified, determine <code>mtx.rows</code> that are below this threshold if $p < 1$ or top $p$ sets if $p > 1$ .
<code>q.limit</code>	A numeric value. If specified, determine <code>mtx.rows</code> that are below this threshold if $q < 1$ or top $q$ sets if $q > 1$ .
<code>p.featt.limit</code>	A numeric value. If specified, determine <code>mtx.rows</code> that are below this threshold if $p.featt < 1$ or top $p.featt$ sets if $p.featt > 1$ (feature p-values).
<code>q.featt.limit</code>	A numeric value. If specified, determine <code>mtx.rows</code> that are below this threshold if $q.featt < 1$ or top $q.featt$ sets if $q.featt > 1$ .
<code>intersect</code>	A logical value. Default is <code>TRUE</code> . If <code>TRUE</code> , use intersection of all specified criteria. If <code>FALSE</code> use union of all specified criteria.
<code>recalc</code>	A logical value. Default is <code>FALSE</code> . If <code>TRUE</code> , recalculate p-values. If <code>FALSE</code> use original set p-values..

**Value**

A list with filtered beam.stats object, updated beam.set.pvals, and filtered beam.featt.pvals.

**Examples**

```
data(beam_stats)
test.pvals <- compute_set_pvalues(beam_stats=beam_stats)
test.featt.pvals <- compute_feature_pvalues(beam_stats=beam_stats)
filt.beam_stats <- subset_beam_result(beam_stats, test.pvals, test.featt.pvals,
                                     endpts=c("EFS", "OS"), q.limit=10, intersect=TRUE,
                                     recalc=FALSE)
```

# Index

## \* datasets

- beam\_dat, [2](#)
- beam\_dat\_sm, [3](#)
- beam\_specs, [4](#)
- beam\_specs\_sm, [4](#)
- beam\_stats, [5](#)
- beam\_stats\_sm, [5](#)
- clinf, [7](#)
- omicann, [14](#)
- omicdat, [14](#)
- setdat, [22](#)
- specs, [23](#)

- beam\_dat, [2](#)
- beam\_dat\_sm, [3](#)
- beam\_specs, [4](#)
- beam\_specs\_sm, [4](#)
- beam\_stats, [5](#)
- beam\_stats\_sm, [5](#)

- check\_beam\_specs, [6](#)
- check\_list\_class, [6](#)
- clean\_Bmtx, [7](#)
- clinf, [7](#)
- compute\_beam\_stats, [8](#)
- compute\_feature\_pvalues, [9](#)
- compute\_set\_pvalues, [9](#)

- extend\_set\_data, [10](#)
- extract\_beam\_stats, [11](#)

- find\_id\_clm, [11](#)

- gen\_beam\_plot\_list, [12](#)
- get\_id\_index, [13](#)

- omicann, [14](#)
- omicdat, [14](#)

- plot\_beam\_boot, [15](#)
- plot\_beam\_clin, [16](#)

- plot\_feat\_clin, [17](#)
- prep\_beam\_data, [18](#)
- prep\_beam\_plot, [19](#)
- prep\_beam\_specs, [20](#)
- print.beam.data, [21](#)
- print.beam.stats, [22](#)

- setdat, [22](#)
- specs, [23](#)
- subset\_beam\_result, [23](#)